

# SensiTek Alk-Phos Anti-Polyvalent (Permanent Red) Stain Kit

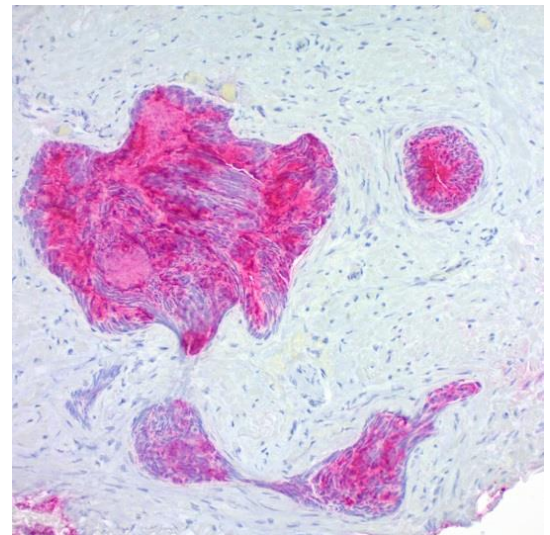
**Description:** The SensiTek staining kit provides unmatched sensitivity with incubation times of 20 minutes each for the Link Antibody and Enzyme Label. This kit includes our Permanent Red chromogen, which can be coverslipped with solvent based mounting media for long term storage.

**Species of Origin:** Goat  
**Antigen Specificity:** Anti-Polyvalent (Mouse, Rat, Rabbit and Guinea Pig).  
**Preadsorbed Against:** Human  
**Enzyme Conjugate:** Alkaline Phosphatase  
**Chromogen Substrate:** Permanent Red

**Uses/Limitations:** Not to be taken internally.  
 For In Vitro Diagnostic Use.  
 Histological applications.  
 Do not use if reagents become cloudy.  
 Do not use past expiration date.  
 Use caution when handling reagents.  
 Non-Sterile.

**Control Tissue:** Any well-fixed tissue section.  
 Frozen tissue section.  
 Cytocentrifuge preparation.

Ordering Information and Current Pricing at [www.scytek.com](http://www.scytek.com)



**Test Capacity:** 70 Slides


Kit Contents:	Item #	Description	Volume
	AAA008	Super Block	8 ml
	ABF008	SensiTek Anti-Polyvalent	8 ml
	ABH008	SensiTek Alk-Phos	8 ml
	PRC002	Permanent Red Concentrate	2 ml
	PRB005	Permanent Red Buffer	5 ml x 8 vials


**Recommended, But Not Included:**

Item #	Description
CPL500	Citrate Plus
HAQ500	Hematoxylin for Automation
BRT500	Bluing Reagent


**Storage:** Store at 2-8°C.

**Precautions:** Avoid contact with skin and eyes.  
 Harmful if swallowed.  
 Follow all Federal, State, and local regulations regarding disposal.

Storage: 2° C  8° C

 ScyTek Laboratories, Inc.  
 205 South 600 West  
 Logan, UT 84321  
 U.S.A.



  
 Emergo Europe  
 Westervoortsedijk 60  
 6827 AT Arnhem, The Netherlands

**Procedure:**

1. Rehydrate tissue slides.
2. In a glass or plastic (Autoclavable) Coplin jar, add 5 ml of Citrate Plus (CPL) and 45 ml of deionized water.
3. Submerge slides in diluted Citrate Plus and loosely cap.
4. Add Distilled water to bottom of Autoclave or Pressure Cooker (about 1 inch deep in Pressure Cooker).
5. Place Coplin jar in Pressure Cooker or Autoclave.
6. Turn heat on and allow pressure to rise to 20-25 PSI.
7. Maintain pressure at 20-25 PSI for 5 minutes.
8. Turn off heat source and allow to cool.
9. When pressure has dropped to ambient, carefully remove lid or open door.
10. Using tongs, remove Coplin Jar and place on counter.
11. Once Coplin Jar cools to room temperature remove slides, rinse several times in buffer and proceed with staining as usual.
12. Rinse in distilled/DI water.
13. Rinse 3 times in buffer.
14. Apply Super Block (blue cap), and incubate for 5 minutes at room temperature to block nonspecific background staining.  
**Note:** Do not exceed 10 minutes or there may be a reduction in desired stain.
15. Rinse 1 time in buffer.
16. Apply primary antibody and incubate according to manufacturer's protocol.
17. Rinse 3 times in buffer.
18. Apply SensiTek Anti-Polyvalent (yellow cap) and incubate for 20 minutes at room temperature.
19. Rinse 2 times in buffer.
20. Apply SensiTek Alk-Phos (red cap) and incubate for 20 minutes at room temperature.
21. Rinse 2 times in buffer.
22. Rinse 1 time in Distilled/DI water.

**Mix Permanent Red Concentrate with Permanent Red Buffer and use immediately.**

23. Add 2 drops (60ul) Permanent Red Chromogen (PRC002) to each 5ml vial of Permanent Red Buffer (PRB005), mix by swirling and apply to tissue for 5 - 10 minutes.
24. Rinse in deionized water.
25. Apply counterstain (suggested: ScyTek HAQ) as desired.
26. Rinse 3 times in distilled water.
27. If counterstained with hematoxylin, apply Bluing Reagent (BRT) and incubate for 5-10 seconds.
28. Rinse immediately in distilled or deionized water.
29. Quickly dehydrate in alcohol and clear in xylene or substitute.  
*Note: Alcohol and Xylene can cause chromogen to leach from tissue over extended periods of time.*
30. Coverslip using a permanent mounting media.

**-Troubleshooting Guide-**Storage:  
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**Overstaining:**

1. Concentration of the primary antibody was too high or the incubation time was too long.
2. Temperature during incubation was too high.
3. Incubation times were too long.

**Non-Specific Background Staining:**

1. Rinsing between steps was inadequate.
2. Tissue was allowed to dry with reagents on.
3. Folds in tissue trapped reagents.
4. Antigen migrated in tissue.
5. Excessive tissue adhesive on slides.
6. Inadequate blocking with protein block.

**Weak Staining:**



1. Primary antibody concentration was too low or incubation time was too short.
2. Reagents are past their expiration date.
3. Inadequate removal of wash buffer between steps, resulting in dilution of reagents.
4. Room temperature was excessively cool.
5. The primary antibody does not recognize an antigen that survives fixation and embedding in high enough amounts.
6. Excessive incubation with protein block (Super Block or normal serum).

**No Staining:**

1. Steps were inadvertently left out.
2. There is no antigen in the tissue.
3. The primary antibody is not of mouse, rat, rabbit or guinea pig origin.
4. Chromogenic substrate has been replaced with another that is not intended for use with alkaline phosphatase.
5. One or more components of the kit have been inactivated.

**References:**

1. Hsieh MH, Jan RL, Wu LS, Chen PC, Kao HF, Kuo WS, Wang JY. Lactobacillus gasseri attenuates allergic airway inflammation through PPAR $\gamma$  activation in dendritic cells. Journal of Molecular Medicine. 2018 Jan 1;96(1):39-51.

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